



KF01007

DPPH Antioxidant Capacity Assay Kit

**96 well plate
100/200/400 tests**

Votre interlocuteur en France, Belgique, Luxembourg et Suisse :
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1. General information

PRECAUTIONS

Please read this manual carefully before beginning the assay.

This product is designed for **research use only**. It is not approved for human or animal use or clinical diagnosis. All chemicals should be handled with care and in accordance with laboratory safety practices. It is recommended to use basic Personal Protective Equipment.

Do not use after the expiration date stated on the packaging.

Do not mix or substitute reagents or materials from other kit batches or vendors.

For the **material safety data sheet** (MSDS) please contact us at info@bioquochem.com

TECHNICAL RECOMMENDATIONS

Store reagents as indicated in **Materials and storage** section.

Be sure to keep the bottle capped when not in use.

Let the components reach room temperature (RT) before use.

Immediately before use, gently invert and rotate reagent bottles several times to mix the contents thoroughly.

Avoid foaming or bubbles when mixing or reconstituting components.

Avoid cross contamination of samples or reagents by changing pipette tips between sample, standard and reagent additions.

Be sure to use the optimal microplate for the assay. Flat bottom transparent microplates for UV/VIS applications, and black microplates for fluorescence measurements.

2. Technical specifications

Available sizes

100/200/400 tests

Required sample volume

20 µL/test

Compatible samples

Biological samples (plasma/serum), foods, beverages and plant extracts

Type of detection

Colorimetric (517 nm)

3. Materials and storage

MATERIALS SUPPLIED

| Item | No. Tests | Units | Storage |
|--------------------------------|-----------|-------|---------|
| Reagent A | 100 | 1 | RT |
| | 200 | 2 | |
| | 400 | 4 | |
| Reagent B | 100 | 1 | 4 °C |
| | 200 | 2 | |
| | 400 | 4 | |
| Standard | 100 | 1 | -20 °C |
| | 200 | 2 | |
| | 400 | 4 | |
| Transparent 96-Well Microplate | 100 | 1 | RT |
| | 200 | 2 | |
| | 400 | 4 | |

MATERIALS NEEDED BUT NOT SUPPLIED

- Double distilled water (ddH₂O) as Milli-Q Ultrapure Water.
- Labware materials (micropipettes, tubes, stirring/mixing equipment).
- Colorimetric microplate reader – equipped with filter for OD 517 nm.

STORAGE CONDITIONS

On receipt, store kit components as indicated above. Under these conditions, the reagents are stable in the original packaging until the expiration date indicated on the outside of the box. After reconstitution, standard solutions are unstable in the presence of oxygen. Prepare a fresh set of standards for every use.

4. Introduction

Antioxidants serve as a protection against the harmful effects of free radical damage. Antioxidant systems include both antioxidative enzymes (superoxide dismutase, catalase, glutathione peroxidase, etc.), and low-molecular weight non-enzymatic compounds (glutathione, uric acid, lipoic acid, bilirubin, coenzyme Q, vitamin C, vitamin A, vitamin E, flavonoids, carotenoids, etc.).

Total antioxidant capacity (TAC) is a global measure of the non-enzymatic antioxidant efficiency that integrates the individual effect of all antioxidants in a given matrix, and their additive, synergistic or antagonistic interactions.

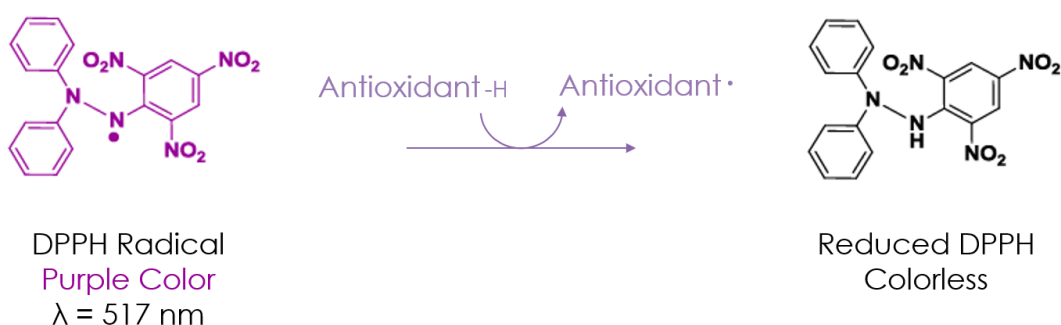
TAC is considered as an important parameter to establish antioxidant status of biological samples. Alterations in the redox status of tissue/organs and body fluids have been linked to several health impairments such as infertility, obesity, cancer, and neurodegenerative diseases.

TAC is also an important tool for plant characterization and for food quality control since the antioxidant levels vary depending on environmental factors, harvesting, aging, storage conditions, etc.

BQC DPPH Assay Kit is an easy, fast, and highly reproducible assay to measure TAC in a wide variety of samples.

5. Assay principle

BQC DPPH TAC Assay Kit is based on the 2,2-diphenyl-1-picrylhydrazyl (DPPH) method. In this method, the DPPH free radical (DPPH[•]) which is a deep purple colored ($\lambda_{\text{max}} = 517 \text{ nm}$) stable organic nitrogen radical, is reduced by antioxidants to the colorless DPPH reduced form. Therefore, the absorbance decrease at 517 nm depends linearly on the antioxidant concentration. The synthetic antioxidant Trolox (included in the kit) is used to standardize the sample TAC relative to Trolox (Trolox Equivalents Antioxidant Capacity, TEAC).



Principle of DPPH Assay Kit

6. Assay preparation

REAGENT PREPARATION

All assay reagents not listed below are ready to use as supplied. Allow the reagents to reach room temperature before use.

R.B. Working Solution: Add 6 mL of Reagent A to each bottle of Reagent B and mix well. Dilute this solution (Solution B) with Reagent A in a 20 mL tube (not provided) to an absorbance of around 1.0 ± 0.2 at 517 nm. This value is usually obtained with a dilution between 1:4 and 1:10 of Solution B with Reagent A. It is recommended to start with the dilution 1:4 (e.g. 5 mL of Solution B with 15 mL of Reagent A) and dilute this solution until the absorbance reaches the desired value.

! CAUTION: R.B. Working Solution must be prepared immediately before use

Standard Solution (Trolox): Add 1 mL of Reagent A to the Standard vial and mix well. Dilute this standard solution 1:10 with Reagent A (e.g. 100 μ L of standard solution with 900 μ L of Reagent A). Use this diluted solution to prepare the standard curve.

STANDARD CALIBRATION

Prepare Trolox (TX) standards for the calibration curve from the 1:10 diluted standard solution according to the following Table. Prepare the standards immediately prior to each assay. Vortex tubes thoroughly. Discard standard solutions after use.

| Standard | Standard Solution 1:10 diluted (μ L) | Reagent A (μ L) | *TEAC (μ M TX) |
|----------|--|----------------------|---------------------|
| Std 1 | 0 | 100 | 0 |
| Std 2 | 10 | 90 | 100 |
| Std 3 | 20 | 80 | 200 |
| Std 4 | 30 | 70 | 300 |
| Std 5 | 40 | 60 | 400 |
| Std 6 | 50 | 50 | 500 |

*Antioxidant activity is expressed as TEAC (Trolox Equivalents Antioxidant Capacity).

PLATE SET UP

BQC recommends running the standards, samples, and blanks at least in duplicate (triplicate recommended). There is no specific pattern for using the wells on the plate. A proposed layout of standards (Std), reagent blank (B) and samples (S) to be measured in duplicate is shown below.

| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
|---|-------|-------|----|----|-----|-----|-----|-----|-----|-----|-----|-----|
| A | Std 1 | Std 1 | S2 | S2 | S10 | S10 | S18 | S18 | S26 | S26 | S34 | S34 |
| B | Std 2 | Std 2 | S3 | S3 | S11 | S11 | S19 | S19 | S27 | S27 | S35 | S35 |
| C | Std 3 | Std 3 | S4 | S4 | S12 | S12 | S20 | S20 | S28 | S28 | S36 | S36 |
| D | Std 4 | Std 4 | S5 | S5 | S13 | S13 | S21 | S21 | S29 | S29 | S37 | S37 |
| E | Std 5 | Std 5 | S6 | S6 | S14 | S14 | S22 | S22 | S30 | S30 | S38 | S38 |
| F | Std 6 | Std 6 | S7 | S7 | S15 | S15 | S23 | S23 | S31 | S31 | S39 | S39 |
| G | B | B | S8 | S8 | S16 | S16 | S24 | S24 | S32 | S32 | S40 | S40 |
| H | S1 | S1 | S9 | S9 | S17 | S17 | S25 | S25 | S33 | S33 | S41 | S41 |

Example of plate layout for the DPPH Assay Kit

7. Sample preparation

The following sample preparation protocols are intended as a guide only. The optimal conditions for sample preparation must be determined by the end user. It is recommended to use fresh samples. If it is not possible, aliquot and store samples appropriately with minimal freeze/thawing.

DPPH Assay Kit can be used to determine TAC in a wide variety of samples like plasma/serum, fruits, vegetables, plants, and beverages.

Biological samples. Serum and plasma samples must be deproteinized (e.g. mix with an equal volume of acetonitrile, incubate 2 minutes and centrifuge at 9500 x g for 10 minutes at 4 °C) before performing the assay.

Food and beverages. Fruit juices and other beverages such as wine, tea, and coffee can be directly measured with appropriate dilutions. If it is required, clarify the sample through filtration prior performing the assay.

For the analysis of other samples like **fruits, vegetables, and plants** an extraction step is usually required. The extraction method varies based upon the sample type. The most common extraction solvents include acid/methanol, acid/ethanol, or acetone.

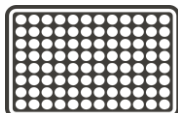
Reagents and materials required for sample preparation are not supplied with the kit. Before doing sample preparation, consider the volume of sample required per test; see **Technical specifications** section.

Make sure that interfering substances present in the sample do not give a significant background. Run proper blanks as necessary (e.g. sample blank should be always evaluated when working with highly colored samples). It is recommended to assay different sample dilutions to ensure the values fall within the standard curve.

8. Assay protocol

Prepare and mix all reagents thoroughly before use. Each standard, sample or blank should be assayed at least in duplicate.

1



Set up the plate design

2



Add **20 µL** of **standard** or **sample** in each well. For **reagent blank** wells add **20 µL** of **Reagent A**

3



- **Standard** and **sample** wells: Add **200 µL** of **R.B. Working Solution**

- **Reagent blank** wells: Add **200 µL** of **Reagent A**

4



Read the **absorbance** of all wells at **517 nm** in end point mode at **RT**

If you need to **adapt this kit** for another form of the assay (for example cuvette), **contact us at** info@bioquochem.com

9. Data analysis

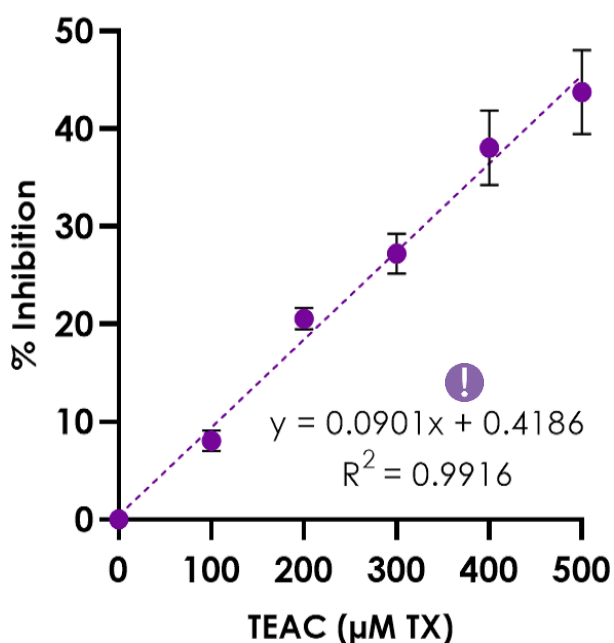
ANALYSIS OF THE STANDARDS

- Calculate the average absorbance of the standards.
- Subtract the average absorbance of the reagent blank from the average absorbance of the standards to obtain the blank-corrected absorbance of the standards.
- Calculate the % inhibition for the standards according to the following equation:

$$\% \text{ Inhibition of Standard} = \left(1 - \frac{A_{\text{Std}}}{A_0}\right) \cdot 100$$

Where A_0 is the blank-corrected absorbance of Std 1 (0 μM TX) and A_{Std} is the blank-corrected absorbance measured for the remaining standards.

- Create a standard curve by plotting the % inhibition of standards as a function of the standard concentration (see **STANDARD CALIBRATION** section). A typical standard curve ($y = \text{slope} \cdot x \pm \text{intercept}$) for this assay is shown below.



TX standard curve with DPPH Assay Kit

- ❗ This standard curve is an example of the data typically obtained with this kit. **DO NOT USE** this standard curve to calculate TAC of your samples. A new standard curve must be performed by the end user.

ANALYSIS OF THE SAMPLES

- Calculate the average absorbance of the samples.
- Subtract the average absorbance of the reagent blank from the average absorbance of each sample to obtain the blank-corrected absorbance of the samples (A_s).
- Calculate the % inhibition for the samples according to the following equation:

$$\% \text{ Inhibition of Sample} = \left(1 - \frac{A_s}{A_0}\right) \cdot 100$$

Where A_0 is the blank-corrected absorbance of Std 1 (0 μM TX) and A_s is the blank-corrected absorbance measured for the sample.

- Calculate the TEAC value of the samples using the following equation. Slope and intercept values are obtained from the standard curve.

$$\text{TEAC } (\mu\text{M TX}) = \left(\frac{\% \text{ Inhibition of Sample} - \text{intercept}}{\text{slope}} \right)$$

When working with diluted samples the concentration values obtained must be multiplied by the dilution factor to obtain the TEAC (μM TX) value of the undiluted sample.

10. Troubleshooting

This troubleshooting table provides potential sources and solutions for common problems observed with BQC Assay Kits. **The problems listed below could occur when using any BQC Assay Kit.** They are not specific for this assay kit.

| Problem | Possible Cause | Recommended Solution |
|--|--|--|
| Wells have color but there is no reading | Plate read at incorrect wavelength | Check the wavelength used in the assay |
| | Incorrect microplate | Use the correct microplate for your application UV/Vis: transparent Fluorescence: black wells/transparent bottom |
| Standard readings do not follow a linear pattern | Pipetting errors in preparation of standards | Avoid pipetting small volumes (<5 µL) Be careful not to splash from well to well |
| | Air bubbles formed in well(s) | Use reverse pipetting technique |
| | Standard stock is at incorrect concentration | Always refer to dilutions described in Assay preparation |
| | Improperly thawed reagents | Thaw all components completely and mix well before use |
| | Use of improperly stored reagents | Store the components appropriately Use fresh components from the standard curve |
| | Incorrect incubation times or temperatures | Refer to Assay protocol |
| Dispersion of standard and sample readings | Pipetting errors | Avoid pipetting small volumes (<5 µL) Be careful not to splash from well to well |
| | Air bubbles formed in well(s) | Use reverse pipetting technique |

| Problem | Possible Cause | Recommended Solution |
|---|---|--|
| Sample erratic values | Samples contain interfering substances | Dilute sample further (if possible) |
| | Inappropriately stored samples or samples used after multiple freeze-thaw cycles | Use fresh samples or store appropriately until use |
| | Samples not deproteinized | Use an appropriate deproteinization protocol |
| | Cells/Tissue samples not homogenized completely | Repeat the sample homogenization |
| | Inappropriate sample dilution buffer | Refer to Assay preparation |
| Sample reading fall outside the detection range | Samples are too diluted/concentrated No analyte/activity is observed in the sample | Re-assay using different sample dilutions |

STILL HAVING PROBLEMS?

Contact BQC if you have any further questions, our team will be pleased to help you:



Phone

+ 34 985 26 92 92



E-mail

info@bioquochem.com



Business hours

Monday-Thursday: 8.30 to 17.00 (CEST)
Friday: 8.00 to 15.00 (CEST)

11. Additional information

BQC DPPH Assay Kit is a quick (< 15 minutes) assay for determining TAC in a wide variety of samples.

Colored compounds with wavelengths similar to DPPH• (e.g. carotenoids and anthocyanins) have been reported to interfere with this assay.

Samples containing proteins may precipitate in the alcoholic reaction medium.

If unexpected results are obtained running your samples, please contact us at info@bioquochem.com

12. Related products

More products available on bioquochem.com

| Reference | Product |
|-----------|-------------------------------------|
| KB03006 | Polyphenol Quantification Assay Kit |
| KB03015 | Anthocyanins Assay Kit |
| KB03033 | NAD/NADH Quantification Assay Kit |

13. Warranties and limitation of liability

BQC shall not in any event be liable for incidental, consequential or special damages of any kind resulting from any use or failure of the products, even if BQC has been advised of the possibility of such damage including, without limitation, liability for loss of use, loss of work in progress, downtime, loss of revenue or profits, failure to realize savings, loss of products of buyer or other use or any liability of buyer to a third party on account of such loss, or for any labor or any other expense, damage or loss occasioned by such product including personal injury or property damage is caused by BQC's gross negligence. Any and all liability of BQC hereunder shall be limited to the amounts paid by the buyer for the product.

Buyer's exclusive remedy and BQC's sole liability hereunder shall be limited to a refund of the purchase price, or the replacement of all material that does not meet our specifications.

Said refund or replacement is conditioned on buyer giving written notice to BQC within 30 days of shipment.

Expiration date: 1 year from the date of fabrication. Expiration date is indicated on the outside of the box.

For further details, please refer to our website bioquochem.com



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