Megazyme

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L-RHAMNOSE

ASSAY PROCEDURE

K-RHAMNOSE 08/18

(*50/100 Manual Assays per Kit) or
(550 Auto-Analyser Assays per Kit) or
(550 Microplate Assays per Kit)

** The number of tests per kit can be doubled if all volumes are halved*

INTRODUCTION:

L-Rhamnose occurs naturally in the L-form and is commonly present as a component of the carbohydrate moiety of eukaryotic glycoproteins and in plant cell wall polysaccharides. The most abundant occurrence of L-rhamnose is within the pectic fraction of plant cell wall polysaccharides particularly as a component of rhamnogalacturonan I, which consists of the repeating disaccharide unit [1,4)-α-D-Gal*p*A-(1,2)-α-L-Rha*p*-(1,4]-, and to a lesser extent in rhamnogalacturonan II.

L-Rhamnose is commonly used as a non-metabolisable marker along with lactulose for dual-permeability testing in the diagnosis of intestinal diseases such as Crohn's disease or coeliac disease.

This kit (**K-RHAMNOSE**) is suitable for the specific measurement of L-rhamnose.

PRINCIPLE:

L-Rhamnose is oxidised by nicotinamide-adenine dinucleotide (NAD^+) in the presence of the enzyme, L-rhamnose dehydrogenase (1).

(1) L-Rhamnose + NAD^+ $\xrightarrow{\text{(L-rhamnose dehydrogenase)}}$ L-rhamno-1,4-lactone + NADH + H^+

The amount of NADH formed in this reaction is stoichiometric with the amount of L-rhamnose. It is the NADH which is measured by the increase in absorbance at 340 nm.

SPECIFICITY, SENSITIVITY, LINEARITY AND PRECISION:

The assay rapidly converts L-rhamnose, and also converts the rare monosaccharide L-lyxose but at approximately 2-fold slower than L-rhamnose. D-Arabinose, L-arabinose, D-galactose, D-glucose, D-fructose, L-fucose, D-mannose and L-xylose do not react.
The smallest differentiating absorbance for the assay is 0.010 absorbance units. This corresponds to 0.6 mg/L of sample solution at the maximum sample volume of 1.00 mL. The detection limit is 1.2 mg/L, which is derived from an absorbance difference of 0.020 with a sample volume of 1.00 mL.

The assay is linear over the range of 5 to 100 µg of L-rhamnose per assay. In duplicate determinations using one sample solution, an absorbance difference of 0.005 to 0.010 may occur. With a sample volume of 1.00 mL, this corresponds to an L-rhamnose concentration of approx. 0.3 to 0.6 mg/L of sample solution. If, in sample preparation, the sample is weighed, e.g. 10 g/L, a difference of 0.02 to 0.05 g/100 g can be expected.

INTERFERENCE:

If the conversion of L-rhamnose has been completed within the time specified in the assay (approx. 5 min), it can be generally concluded that no interference has occurred. However, this can be further checked by adding L-rhamnose (approx. 50 µg in 0.1 mL) to the cuvette on completion of the reaction. A significant increase in the absorbance should be observed.

Interfering substances in the sample being analysed can be identified by including an internal standard. Quantitative recovery of this standard would be expected. Losses in sample handling and extraction are identified by performing recovery experiments, i.e. by adding L-rhamnose to the sample in the initial extraction steps.

SAFETY:

The general safety measures that apply to all chemical substances should be adhered to.

For more information regarding the safe usage and handling of this product please refer to the associated SDS that is available from the Megazyme website.

KITS:

Kits suitable for performing 50/100 assays in manual format (or 550 assays in auto-analyser format or microplate format) are available from Megazyme. The kits contain the full assay method plus:

Bottle 1: Buffer (9 mL, pH 10) plus sodium azide (0.02% w/v) as a preservative.
Stable for > 2 years at 4°C.

Bottle 2: NAD^+ plus PVP. Freeze dried powder.
Stable for > 5 years below -10°C.

Bottle 3: L-Rhamnose dehydrogenase solution (2.75 mL).
Stable for > 4 years below -10°C.

Bottle 4: L-Rhamnose (5 mL; 0.5 mg/mL).
Stable for > 2 years at 4°C.

PREPARATION OF REAGENT SOLUTIONS (SUPPLIED):

1. Use the contents of bottle 1 as supplied.
   Stable for > 2 years at 4°C.
2. Dissolve the contents of bottle 2 in 6 mL of distilled water. **Stable for > 1 year at 4°C** or stable for > 2 years below -10°C (to avoid repetitive freeze/thaw cycles, divide into appropriately sized aliquots and store in polypropylene tubes).

3. Use the contents of bottle 3 as supplied. Before opening for the first time, shake the bottle to remove any enzyme that may have settled on the rubber stopper. Subsequently store the bottle in an upright position. **Swirl the bottle to mix contents before use**.
   Stable for > 4 years below -10°C.
4. Use the contents of bottle 4 as supplied.
   Stable for > 2 years at 4°C.

NOTE: The L-rhamnose standard solution (bottle 4) is only assayed where there is some doubt about the accuracy of the spectrophotometer being used or where it is suspected that inhibition is being caused by substances in the sample. To test the assay using the standard solution, add 0.1 mL (50 µg L-rhamnose) of standard solution to the assay cuvette in place of the sample. The concentration of L-rhamnose is determined directly from the extinction coefficient of NADH (page 5).

EQUIPMENT (RECOMMENDED):

1. Disposable plastic cuvettes (1 cm light path, 3 mL).
2. Micro-pipettors, e.g. Gilson Pipetman® (20 µL, 200 µL and 1 mL).
3. Positive displacement pipettor, e.g. Eppendorf Multipette®
   - with 5 mL Combitip® [to dispense 0.1 mL aliquots of buffer (solution 1) and 0.1 mL aliquots of NAD^+/PVP (solution 2)].
4. Stop clock.
5. Analytical balance.
6. Spectrophotometer set at 340 nm.
7. Vortex mixer (e.g. IKA® Yellowline Test Tube Shaker TTS2).

A. MANUAL ASSAY PROCEDURE:

NOTE: The standard Manual Assay Procedure provides 50 assays per kit. To obtain 100 assays per kit, reduce the volumes of the assay components stated in the procedure table (below) by half.

Wavelength: 340 nm
Cuvette: 1 cm light path (glass or plastic)
Temperature: ~ 25°C
Final volume: 2.35 mL
Sample solution: 5-100 µg of L-rhamnose per cuvette (in 0.1-1.0 mL sample volume)

Read against air (without a cuvette in the light path) or against water

Pipette into cuvettes	Blank	Sample
distilled water (at ~ 25°C)	2.10 mL	2.00 mL
sample	-	0.10 mL
solution 1 (buffer)	0.10 mL	0.10 mL
solution 2 (NAD^+)	0.10 mL	0.10 mL
Mix*, read the absorbances of the solutions (A_1) after approx. 3 min and start the reactions by addition of:		
solution 3 (L-RDH)	0.05 mL	0.05 mL
Mix* and read the absorbances of the solutions (A_2) at the end of the reaction (approx. 5 min). If the reaction has not stopped after 5 min, continue to read the absorbances at 1 min intervals until the absorbances increase constantly over 1 min**.		

* for example with a plastic spatula or by gentle inversion after sealing the cuvette with a cuvette cap or Parafilm®.

** if this "creep" rate is greater for the sample than for the blank, extrapolate the sample absorbances back to the time of addition of solution 3.

CALCULATION:

Determine the absorbance difference (A_2-A_1) for both blank and sample. Subtract the absorbance difference of the blank from the absorbance difference of the sample, thereby obtaining $\Delta A_{L\text{-rhamnose}}$. The value of $\Delta A_{L\text{-rhamnose}}$ should as a rule be at least 0.100 absorbance units to achieve sufficiently accurate results.

The concentration of L-rhamnose can be calculated as follows:

$$c = \frac{V \times MW}{\varepsilon \times d \times v} \times \Delta A_{L\text{-rhamnose}} \quad [g/L]$$

where:

V = final volume [mL]
MW = molecular weight of L-rhamnose [g/mol]
ε = extinction coefficient of NADH at 340 nm
= 6300 [l x mol^{-1} x cm^{-1}]
d = light path [cm]
v = sample volume [mL]

It follows for L-rhamnose:

$$c = \frac{2.35 \times 164.16}{6300 \times 1.0 \times 0.1} \times \Delta A_{L\text{-rhamnose}} \quad [g/L]$$

$$= 0.6123 \times \Delta A_{L\text{-rhamnose}} \quad [g/L]$$

If the sample has been diluted during preparation, the result must be multiplied by the dilution factor, F.

When analysing solid and semi-solid samples which are weighed out for sample preparation, the content (g/100 g) is calculated from the amount weighed as follows:

Content of L-rhamnose:

$$= \frac{c_{L\text{-rhamnose}}\ [g/L\ \text{sample solution}]}{\text{weight}_{sample}\ [g/L\ \text{sample solution}]} \times 100 \quad [g/100\ g]$$

NOTE: These calculations can be simplified by using the Megazyme ***Mega-Calc***™, downloadable from where the product appears in the Megazyme web site (www.megazyme.com).

B. AUTO-ANALYSER ASSAY PROCEDURE:

NOTES:

1. The Auto-Analyser Assay Procedure for L-rhamnose can be performed using either a single point standard or a full calibration curve.
2. For each batch of samples that is applied to the determination of L-rhamnose **either a single point standard or a calibration curve must be performed concurrently using the same batch of reagents.**

Reagent preparation is performed as follows:

Preparation of R1:

Component	Volume
distilled water solution 1 (buffer) solution 2 (NAD^+/PVP)	54 mL 3 mL 3 mL (after adding 6 mL of H_2O to bottle 2)
Total volume	60 mL

Preparation of R2:

Component	Volume
distilled water solution 1 (buffer) solution 3 (L-RDH)	3.2 mL 1.0 mL 1.3 mL
Total volume	5.5 mL

EXAMPLE METHOD:

R1:	0.200 mL
Sample:	~ 0.01 mL
R2:	0.02 mL
Reaction time:	~ 5 min at 25°C or ~ 4 min at 37°C
Wavelength:	340 nm
Prepared reagent stability:	> 14 days at 4°C / > 1 year below -10°C
Calculation:	endpoint
Reaction direction:	increase
Linearity:	up to 0.9 g/L of L-rhamnose using 0.01 mL sample volume

C. MICROPLATE ASSAY PROCEDURE:

NOTES:

1. The Microplate Assay Procedure for L-rhamnose can be performed using either a single point standard or a full calibration curve.
2. For each batch of samples that is applied to the determination of L-rhamnose **either a single point standard or a calibration curve must be performed concurrently using the same batch of reagents.**

Wavelength: 340 nm
Microplate: 96-well (e.g. clear flat-bottomed, glass or plastic)
Temperature: ~ 25°C
Final volume: 0.235 mL
Linearity: 0.1-10 µg of L-rhamnose per well (in 0.01-0.20 mL sample volume)

Pipette into wells	Blank	Sample	Standard
distilled water	0.210 mL	0.200 mL	0.200 mL
sample solution	-	0.010 mL	-
standard solution	-	-	0.010 mL
solution 1 (buffer)	0.010 mL	0.010 mL	0.010 mL
solution 2 (NAD^+/PVP)	0.010 mL	0.010 mL	0.010 mL
Mix*, read the absorbances of the solutions (A_1) after approx. 3 min and start the reactions by addition of:			
solution 3 (L-RDH)	0.005 mL	0.005 mL	0.005 mL
Mix* and read the absorbances of the solutions (A_2) at the end of the reaction (approx. 5 min). If the reaction has not stopped after 5 min, continue to read the absorbances at 1 min intervals until the absorbances increase constantly over 1 min**.			

* for example using microplate shaker, shake function on a microplate reader or repeated aspiration (e.g. using a pipettor set at 50-100 µL volume).

** if this "creep" rate is greater for the sample than for the blank, extrapolate the sample absorbances back to the time of addition of suspension 3.

CALCULATION (Microplate Assay Procedure):

$$g/L = \frac{\Delta A_{sample}}{\Delta A_{standard}} \times g/L\ standard \times F$$

If the sample is diluted during preparation, the result must be multiplied by the dilution factor, F.

SAMPLE PREPARATION:

1. Sample dilution.

For the manual assay format the amount of L-rhamnose present in the cuvette (i.e. in the 0.1 mL of sample being analysed) should range between 5 and 100 µg. The sample solution must therefore be diluted sufficiently to yield a concentration between 0.05 and 1.00 g/L.

Dilution Table

Estimated concentration of L-rhamnose (g/L)	Dilution with water	Dilution factor (F)
< 1.00	No dilution required	1
1.00-10.0	1 + 9	10
10.0-100	1 + 99	100
> 100	1 + 999	1000

If the value of $\Delta A_{\text{L-rhamnose}}$ is too low (e.g. < 0.100), weigh out more sample or dilute less strongly. Alternatively, the sample volume to be pipetted into the cuvette can be increased up to 2.1 mL, making sure that the sum of the sample and distilled water components in the reaction is 2.1 mL and using the new sample volume in the equation.

2. Sample clarification.

a. Solutions:

Carrez I solution. Dissolve 3.60 g of potassium hexacyanoferrate (II) $\{K_4[Fe(CN)_6].3H_2O\}$ (Sigma cat. no. P9387) in 100 mL of distilled water. Store at room temperature.

Carrez II solution. Dissolve 7.20 g of zinc sulphate ($ZnSO_4.7H_2O$) (Sigma cat. no. Z4750) in 100 mL of distilled water. Store at room temperature.

Sodium hydroxide (NaOH, 100 mM). Dissolve 4 g of NaOH in 1 L of distilled water. Store at room temperature.

b. Procedure:

Pipette the liquid sample into a 100 mL volumetric flask which contains approx. 60 mL of distilled water, or weigh sufficient quantity of the sample into a 100 mL volumetric flask and add 60 mL of distilled water. Carefully add 5 mL of Carrez I solution, 5 mL of Carrez II solution and 10 mL of NaOH solution (100 mM). Mix after each addition. Fill the volumetric flask to the mark, mix and filter.

3. General considerations.

(a) Liquid samples: clear, slightly coloured and approximately neutral, liquid samples can be used directly in the assay.

(b) Acidic samples: if > 0.1 mL of an acidic sample is to be used undiluted (such as wine or fruit juice), the pH of the solution should be increased to approx. 7.4 using 2 M NaOH, and the solution incubated at room temperature for 30 min.

(c) Carbon dioxide: samples containing a significant amount of carbon dioxide should be degassed by increasing the pH to approx. 7.4 with 2 M NaOH and gentle stirring, or by stirring with a glass rod.

(d) Coloured samples: an additional sample blank, i.e. sample with no L-rhamnose dehydrogenase, may be necessary in the case of coloured samples.

(e) Strongly coloured samples: if used undiluted, strongly coloured samples should be treated by the addition of 0.2 g of polyvinylpyrrolidone (PVPP)/10 mL of sample. Shake the tube vigorously for 5 min and then filter through Whatman No. 1 filter paper.

(f) Solid samples: homogenise or crush solid samples in distilled water and filter if necessary.

(g) Samples containing fat: extract such samples with hot water at a temperature above the melting point of the fat, e.g. in a 100 mL volumetric flask. Adjust to room temperature and fill the volumetric flask to the mark with distilled water. Store on ice or in a refrigerator for 15-30 min and then filter. Discard the first few mL of filtrate and use the clear supernatant (which may be slightly opalescent) for assay. Alternatively, clarify with Carrez reagents.

(h) Samples containing protein: deproteinise samples containing protein with Carrez reagents.

SAMPLE PREPARATION EXAMPLES:

(a) Determination of free L-rhamnose in plant samples.

Mill plant materials to pass a 0.5 mm screen. Weigh out 1.0 g of sample and extract with 90 mL of water (heated to 80°C). Quantitatively transfer to a volumetric flask and dilute to the mark with distilled water. Mix, filter and use the appropriately diluted clear solution for the assay.

(b) Determination of L-rhamnose in fermentation samples and cell culture medium.

Incubate an aliquot (approx. 10 mL) of the solution at approx. 90-95°C for 10 min to inactivate enzyme activity. Centrifuge or filter and use the supernatant or clear filtrate (diluted according to the dilution table, if necessary) for the assay. Alternatively, deproteinisation can be performed with Carrez reagents. Homogenise gelatinous agar media with water and treat further as described above.

(c) Determination of L-rhamnose in polysaccharides and fibrous plant material.

Mill plant material or polysaccharide to pass a 0.5 mm screen using a Retsch centrifugal mill, or similar. Accurately weigh approx. 100 mg of material into a Corning screw-cap culture tube (16 x 125 mm). Add 5 mL of 1.3 M HCl to each tube and cap the tubes. Incubate the tubes at 100°C for 1 h. Stir the tubes intermittently during the incubation. Cool the tubes to room temperature, carefully loosen the caps and add 5 mL of 1.3 M NaOH. Quantitatively transfer the contents of the tube to a 100 mL volumetric flask using distilled water and adjust the volume to 100 mL with distilled water. Mix thoroughly by inversion and filter an aliquot of the solution through Whatman No. 1 filter paper or centrifuge at 1,500 *g* for 10 min. *Typically, no further dilution is required and a sample volume of 0.05-2.1 mL is satisfactory.*

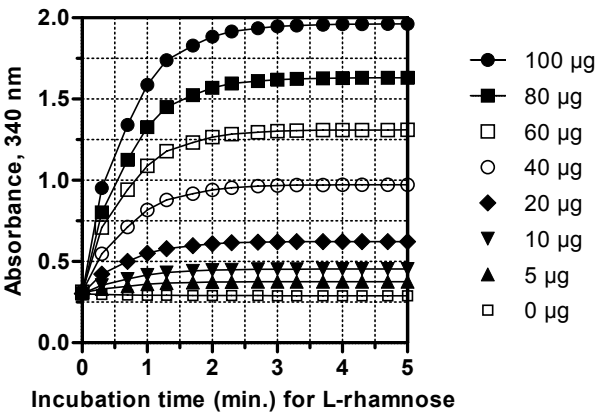

Figure 1. Increase in absorbance at 340 nm on incubation of 0-100 µg of L-rhamnose with L-rhamnose dehydrogenase in the presence of NAD^+ at 25°C using 1 cm path-length cuvettes (Manual Format; page 4).

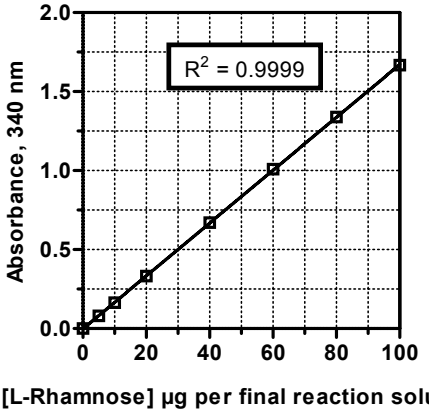

Figure 2. Calibration curve showing the linearity of **K-RHAMNOSE** from 0-100 µg of L-rhamnose. The reactions used to generate this calibration curve were performed at 25°C using 1 cm path-length cuvettes (Manual Format; page 4).

Megazyme

Bray Business Park, Bray,
Co. Wicklow,
IRELAND.

Telephone: (353.1) 286 1220
Facsimile: (353.1) 286 1264
Internet: www.megazyme.com
E-Mail: info@megazyme.com
